Folate Microbiologic Assay Training Poster

Part 2: Set up and Dilute Samples



Keep a run sheet to make note of the medium lot#, calibrator lot#, sample dilution factor, or any other specific information.



While samples are thawing in a 12x4 rack, add QC and blank samples to the rack.



Use a parallel setup for sample vials and glass tubes that contain the diluted samples. Label the glass tubes by 1, 2, 3..., and arrange them in a separate 12x6 rack. One 12x6 rack holds 48 samples, which fit into two 96-well plates.



An experienced analyst can prepare five 96-well plates per run: one calibrator plate and four sample plates. Each sample plate contains one QC sample, 22 unknowns and one blank sample (negative control).



For regular dilution of serum samples (~1:100) or whole blood lysates (~1:140), add 1,475 μ L or 2,075 μ L of 0.5% sodium ascorbate, respectively into each glass tube using a 25-mL automated repeater pipette.



Thoroughly vortex-mix each sample vial before transferring 15 μ L of sample into the glass tubes containing the appropriate amount of 0.5% sodium ascorbate using a single channel manual pipette (10-100 μ L range). Ensure that there are no air bubbles in the pipette tip.



When completed, cap the glass tubes; keep them on the bench protected from light. Place the 12x4 rack with the sample vials in the refrigerator until they are scanned (do not change sequence of sample vials).



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