- Benzarti E, Linden A, Desmecht D, Garigliany M. Mosquito-borne epornitic flaviviruses: an update and review. J Gen Virol. 2019;100:119–32. https://doi. org/10.1099/jgv.0.001203
- Cadar D, Lühken R, van der Jeugd H, Garigliany M, Ziegler U, Keller M, et al. Widespread activity of multiple lineages of Usutu virus, western Europe, 2016. Euro Surveill. 2017;22:30452. https://doi.org/10.2807/ 1560-7917.ES.2017.22.4.30452
- Oude Munnink BB, Münger E, Nieuwenhuijse DF, Kohl R, van der Linden A, Schapendonk CME, et al. Genomic monitoring to understand the emergence and spread of Usutu virus in the Netherlands, 2016–2018. Sci Rep. 2020;10:2798. https://doi.org/10.1038/s41598-020-59692-y
- Aberle SW, Kolodziejek J, Jungbauer C, Stiasny K, Aberle JH, Zoufaly A, et al. Increase in human West Nile and Usutu virus infections, Austria, 2018. Euro Surveill. 2018;23:23. https://doi.org/10.2807/1560-7917. ES.2018.23.43.1800545
- Vilibic-Cavlek T, Petrovic T, Savic V, Barbic L, Tabain I, Stevanovic V, et al. Epidemiology of Usutu virus: the European scenario. Pathogens. 2020;9:E699. https://doi.org/ 10.3390/pathogens9090699
- Hönig V, Palus M, Kaspar T, Zemanova M, Majerova K, Hofmannova L, et al. Multiple lineages of Usutu virus (Flaviviridae, Flavivirus) in blackbirds (Turdus merula) and mosquitoes (Culex pipiens, Cx. modestus) in the

Czech Republic (2016–2019). Microorganisms. 2019;7:568. https://doi.org/10.3390/microorganisms7110568

- Folly AJ, Lawson B, Lean FZ, McCracken F, Spiro S, John SK, et al. Detection of Usutu virus infection in wild birds in the United Kingdom, 2020. Euro Surveill. 2020;25:2001732. https://doi.org/10.2807/1560-7917. ES.2020.25.41.2001732
- Rubel F, Brugger K, Hantel M, Chvala-Mannsberger S, Bakonyi T, Weissenböck H, et al. Explaining Usutu virus dynamics in Austria: model development and calibration. Prev Vet Med. 2008;85:166–86. https://doi.org/10.1016/ j.prevetmed.2008.01.006
- Zannoli S, Sambri V. West Nile virus and Usutu virus co-circulation in Europe: epidemiology and implications. Microorganisms. 2019;7:E184. https://doi.org/10.3390/ microorganisms7070184
- Fros JJ, Miesen P, Vogels CB, Gaibani P, Sambri V, Martina BE, et al. Comparative Usutu and West Nile virus transmission potential by local Culex pipiens mosquitoes in northwestern Europe. One Health. 2015;1:31–6. https://doi.org/10.1016/j.onehlt.2015.08.002

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COMMENT LETTERS

Guillain-Barré Syndrome Associated with COVID-19 Vaccination

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To the Editor: With interest we read the article by Shao et al. (1) about the frequency of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccination-associated Guillain-Barré syndrome (SCo-VaG) among 18,269 healthcare workers in Taiwan who had received the AstraZeneca vaccine (AZV; https://www.astrazeneca.com). Only 1 vaccinee experienced SCoVaG during the study period (1). The study is appealing but raises concerns.

Recently, our review of 19 SCoVaG patients, for whom data were collected through June 2021, was published (2). The 9 men and 10 women in the study were 20-86 years of age. All patients experienced SCoVaG after the first vaccine dose. AZV was given to 14 patients, the Pfizer-BioNTech (https://www. pfizer.com) vaccine to 4 patients, and the Johnson & Johnson (https://www.jnj.com) vaccine to 1 patient. Latency between vaccination and SCoVaG onset ranged from 3 hours to 39 days. Patients received intravenous immune globulin (n = 13), steroids (n = 3), or no therapy (n = 3). Six patients required mechanical ventilation. One patient recovered completely; 9 achieved partial recovery (2). Only 1 of the studies included in our review mentioned the total number of vaccinated persons (3); in that study, 7 persons among 1.2 million vaccinated persons were found to have SCoVaG (3).

In addition, data on 389 patients with SCoVaG were collected in a recent review about the neurologic adverse events of SARS-CoV-2 vaccination (4). However, no individual data were provided for 337 of these patients (4). Among the 53 patients for whom individual data were available, AZV was given to 39 patients, Pfizer-BioNTech vaccine to 9 patients, and Johnson & Johnson vaccine to 2 patients.

For the Shao et al. report (1), we wondered why the oldest healthcare worker was 86 years of age. Also missing were the specific treatment and outcome of the patient with SCoVaG.

Available data suggest that SCoVaG is a rare complication of SARS-CoV-2 vaccination, irrespective of the vaccine brand used. SCoVaG should be diagnosed early so treatment can be initiated promptly. Whether the beneficial effect of SARS-CoV-2 vaccination outweighs the risk for adverse events (e.g., Guillain-Barré syndrome) remains a matter of discussion (5).

References

- 1. Shao SC, Wang CH, Chang KC, Hung MJ, Chen HY, Liao SC. Guillain-Barré syndrome associated with COVID-19 vaccination. Emerg Infect Dis. 2021;27:3175–8. https://doi.org/10.3201/eid2712.211634
- Finsterer J, Scorza FA, Scorza CA. Post SARS-CoV-2 vaccination Guillain-Barre syndrome in 19 patients. Clinics (São Paulo). 2021;76:e3286. https://doi.org/10.6061/ clinics/2021/e3286
- Maramattom BV, Krishnan P, Paul R, Padmanabhan S, Cherukudal Vishnu Nampoothiri S, Syed AA, et al. Guillain-Barré syndrome following ChAdOx1-S/nCoV-19 vaccine. Ann Neurol. 2021;90:312–4. https://doi.org/10.1002/ ana.26143
- Finsterer J. Neurological side effects of SARS-CoV-2 vaccinations. Acta Neurol Scand. 2021. https://doi.org/ 10.1111/ane.13451
- Koike H, Chiba A, Katsuno M. Emerging infection, vaccination, and Guillain-Barré syndrome: a review. Neurol Ther. 2021;10:523–37. https://doi.org/10.1007/ s40120-021-00261-4

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SARS-CoV-2 Cross-Reactivity in Prepandemic Serum from Rural Malaria-Infected Persons, Cambodia

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To the Editor: We read with interest the observations by Manning et al. (1) that serum collected from malaria-infected persons in Cambodia before the coronavirus disease (COVID-19) pandemic harbored seroreactivity against severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) antigens but lacked neutralizing activity. These results suggest that malaria exposure may increase background reactivity in SARS-CoV-2 serosurveys and more specific measures of exposure, such as surrogate virus neutralization tests (sVNTs), may be necessary to capture functional SARS-CoV-2 seroreactivity in malaria-endemic areas. Additional studies in settings with distinct malaria transmission intensities would generalize and strengthen these findings.

One hypothesis for the unexpectedly moderate burden of SARS-CoV-2 in malaria-endemic countries in Africa is that exposure to Plasmodium falciparum confers functional protection against COVID-19 through cross-reactivity or general immune activation. To test this hypothesis, we analyzed 237 dried blood spot samples taken in January 2020 (prepandemic) from P. falciparum-exposed persons in a high-transmission setting in western Kenya for the presence of SARS-CoV-2 neutralizing antibodies (nAbs) using the GenScript SARS-CoV2 sVNT assay (https://www.genscript. com). Monthly P. falciparum real-time PCR results were collected in a previous study (2) for 138/237 persons in the 12 months prior to January 2020. Of these, 131 (95%) were infected with *P. falciparum* at least 1 time in 2019, suggesting that most persons included in this screening had been recently exposed to malaria parasites.

Consistent with findings in Manning et al. (1), none of the 237 people harbored SARS-CoV-2 nAbs, despite high prior levels of exposure to *P. falciparum*. Although nAbs are subject to decay after infection (3), this lack of nAb activity suggests that sVNTs offer a more specific measure of SARS-CoV-2 exposure than standard ELISAs (4). We further suggest that, given that protection from SARS-CoV-2 infection may be associated with the presence of nAbs (5), their absence in samples from both the Manning et al. study