

Prevention and Control (5). Similar to our findings, a study from Denmark reported 47 Omicron BA.2 reinfections that occurred 20–60 days after a primary BA.1 infection (M. Stegger et al., unpub. data, <https://www.medrxiv.org/content/10.1101/2022.02.19.22271112v1>).

The first limitation of our study is that the number of cases was small. Second, we cannot exclude that some cases might have been concurrent infections with different subvariants, notably in the 3 cases that had a 7-day interval between the detection of 2 subvariants. In Marseille, the short time between emergence of different Omicron subvariants might have favored co-infections with different subvariants circulating within the population (Figure). Co-infections can be missed if the quantitative PCR has inadequate sensitivity, and whole-genome sequencing might fail to detect a variant with low prevalence in a patient. Finally, because most reinfection cases were identified from samples transferred to our laboratory by external entities, we were unable to describe COVID-19 vaccination and clinical status of the patients. Nonetheless, our results suggest that the currently used definitions for SARS-CoV-2 reinfection require revision with regard to the duration between primary and secondary infections.

This work was supported by the “Investments for the Future” program managed by the National Agency for Research (ANR) (Méditerranée-Infection 10-IAHU-03), Région Provence Alpes Côte d’Azur, European funding agency FEDER PRIMMI (Fonds Européen de Développement Régional-Plateformes de Recherche et d’Innovation Mutualisées Méditerranée Infection) (FEDER PA 0000320 PRIMMI), Ministry of Higher Education, Research and Innovation (Ministère de l’Enseignement supérieur, de la Recherche et de l’Innovation), and Ministry of Solidarity and Health (Ministère des Solidarités et de la Santé), France.

About the Author

Dr. Nguyen is a medical doctor and PhD student at the Aix Marseille University and Institut Hospitalo-Universitaire Méditerranée Infection, Marseille, France. Her research interests focus on long COVID and SARS-CoV-2 reinfections.

References

1. Nevejan L, Cuypers L, Laenen L, Van Loo L, Vermeulen F, Wollants E, et al. Early SARS-CoV-2 reinfections within 60 days and implications for retesting policies. *Emerg Infect Dis*. 2022;28:1729–31. <https://doi.org/10.3201/eid2808.220617>
2. Nguyen NN, Houhamdi L, Hoang VT, Stoupan D, Fournier PE, Raoult D, et al. High rate of reinfection with

the SARS-CoV-2 Omicron variant. *J Infect*. 2022;85:174–211. <https://doi.org/10.1016/j.jinf.2022.04.034>

3. Colson P, Fournier PE, Chaudet H, Delerce J, Giraud-Gatineau A, Houhamdi L, et al. Analysis of SARS-CoV-2 variants from 24,181 patients exemplifies the role of globalization and zoonosis in pandemics. *Front Microbiol*. 2022;12:786233. <https://doi.org/10.3389/fmicb.2021.786233>
4. Centers for Disease Control and Prevention. Coronavirus disease 2019 (COVID-19) 2021 case definition [cited 2022 Sep 8]. <https://ndc.services.cdc.gov/case-definitions/coronavirus-disease-2019-2021>
5. European Centre for Disease Prevention and Control. Reinfection with SARS-CoV-2: implementation of a surveillance case definition within the EU/EEA [cited 2022 Sep 8]. <https://www.ecdc.europa.eu/en/publications-data/reinfection-sars-cov-2-implementation-surveillance-case-definition-within-eueea>

Address for correspondence: Philippe Gautret, VITROME, Institut Hospitalo-Universitaire Méditerranée Infection, 19-21 Boulevard Jean Moulin, 13385 Marseille CEDEX 05, France; email: philippe.gautret@club-internet.fr

Human Parainfluenza Virus in Homeless Shelters before and during the COVID-19 Pandemic, Washington, USA

Eric J. Chow, Amanda M. Casto, Reigran Sampoleo, Margaret G. Mills, Peter D. Han, Hong Xie, Brian Pfau, Tien V. Nguyen, Jaydee Sereewit, Julia H. Rogers, Sarah N. Cox, Melissa A. Rolfes, Constance Ogokeh, Emily Mosites, Timothy M. Uyeki, Alexander L. Greninger, James P. Hughes, M. Mia Shim, Nancy Sugg, Jeffrey S. Duchin, Lea M. Starita, Janet A. Englund, Pavitra Roychoudhury, Helen Y. Chu

Author affiliations: University of Washington, Seattle, Washington, USA (E.J. Chow, A.M. Casto, R. Sampoleo, M.G. Mills, P.D. Han, H. Xie, B. Pfau, T.V. Nguyen, J. Sereewit, J.H. Rogers, S.N. Cox, A.L. Greninger, J.P. Hughes, M.M. Shim, N. Sugg, J.S. Duchin, L.M. Starita, P. Roychoudhury, H.Y. Chu); Fred Hutchinson Cancer Research Center, Seattle (A.M. Casto, A.L. Greninger, J.P. Hughes, P. Roychoudhury); Brotman Baty Institute for

Precision Medicine, Seattle (P.D. Han, H. Xie, B. Pfau, L.M. Starita); Centers for Disease Control and Prevention, Atlanta, Georgia, USA (M.A. Rolfes, C. Ogokeh, E. Mosites, T.M. Uyeki); Military and Health Research Foundation, Laurel, Maryland, USA (C. Ogokeh); Public Health Seattle and King County, Seattle (M.M. Shim, J.S. Duchin); University of Washington Seattle Children’s Research Institute, Seattle (J.A. Englund)

DOI: <https://doi.org/10.3201/eid2811.221156>

To determine the epidemiology of human parainfluenza virus in homeless shelters during the COVID-19 pandemic, we analyzed data and sequences from respiratory specimens collected in 23 shelters in Washington, USA, during 2019–2021. Two clusters in children were genetically similar by shelter of origin. Shelter-specific interventions are needed to reduce these infections.

Human parainfluenza virus (HPIV) contributes to acute respiratory tract infection burden in young children (1) and adults (2). Persons experiencing homelessness are among those at risk for respiratory viral complications caused by chronic disease burden, mental illness, and social inequities. Homeless shelters might lack resources to reduce viral transmission by using nonpharmaceutical interventions (NPIs). We describe HPIV epidemiology in homeless shelters in King County, Washington, USA, before and during the COVID-19 pandemic.

We analyzed respiratory virus surveillance data from 2 previously described homeless shelter studies (3,4) conducted during October 2019–May 2021. Eligible participants were residents at 1 of 23 homeless shelters who were ≥3 months of age and had a cough or ≥2 other acute respiratory illness symptoms. At enrollment, consenting participants or guardians completed questionnaires, and upper respiratory

specimens were collected; each enrollment was considered 1 encounter. Once a month, persons were eligible to enroll, regardless of symptoms. Beginning April 1, 2020, enrollment expanded to residents and staff, regardless of symptoms. Participants could enroll multiple times; encounters were linked by name and birthdate.

We tested samples by using a TaqMan reverse transcription PCR platform that included influenza virus (A, B, C), respiratory syncytial virus, HPIV (1–4), human coronaviruses, rhinovirus, enterovirus, human bocavirus, human parechovirus, human metapneumovirus, adenovirus, and SARS-CoV-2 (beginning January 1, 2020). A cycle threshold value was generated. We typed HPIV-positive specimens, performed whole-genome sequencing by using hybrid capture on specimens that had a cycle threshold value <22 (Appendix, <https://wwwnc.cdc.gov/EID/article/28/11/22-1156-App1.pdf>), and submitted genomes to GenBank (Appendix Table 1). We aligned shelter consensus genomes generated with corresponding HPIV type genomes from GenBank, generated type-specific phylogenetic trees, and visualized trees by using NextStrain Auspice software (<https://github.com>). We analyzed the data descriptively by using SAS software version 9.4 (<https://www.sas.com>).

During October 2019–May 2021, the study conducted 14,464 encounters with 3,281 unique participants (median age 37 years, range 0.3–85 years; 16% children; 17% shelter staff) (Appendix Figure 1). Among 1,569 encounters with positive virus test results, 32 (2%) encounters from 29 unique participants were HPIV positive (median age 29 years, range 0.3–64 years; 62% children, 45% female, 52% white, 100% resident; 10% had ≥1 chronic condition) (Appendix Table 2). Most HPIV-positive encounters

Table. Human parainfluenza virus detection across 23 homeless shelters, King County, Washington, USA, October 2019–May 2021*

Time period	Type of shelter	Total	Human parainfluenza virus, no. (%) positive	Human parainfluenza virus types
Before April 1, 2020	Shelters: family (sites D, E, O)	303	16 (5.3)	HPIV-1, n = 5; HPIV-3, n = 6; HPIV-4, n = 5; HPIV-1, n = 1
	Shelters: adults 18–25 y (site C)	89	1 (1.1)	
	Shelters: adults ≥18 y (sites A, B, F, L)	845	3 (0.4)	HPIV-1, n = 2; HPIV untyped, n = 1
After April 1, 2020	Shelters: adults ≥50 y (site M)	453	3 (0.7)	HPIV-1, n = 2; HPIV untyped, n = 1
	Shelters: family (sites: D, E, H, N, O, OF, OG)	4,764	8 (0.2)	HPIV-3, n = 5; HPIV untyped, n = 3
	Shelters: adults 18–25 y (sites C, OH)	1,228	0	NA
	Shelters: adults ≥18 y (sites A, B, F, G, J, K, L, OB, OD)	6,078	1 (0.02)	HPIV untyped, n = 1
	Shelters: adults ≥50 y (sites I, M, OA, OC, OE)	661	0	NA
Total		14,421†	32 (0.2)	HPIV-1, n = 10; HPIV-3, n = 11; HPIV-4, n = 5; HPIV untyped, n = 6

*A Washington State Stay-At-Home ordinance we issued on March 23, 2020. HPIV, human parainfluenza virus; NA, not available.

†n = 43 encounters for which dates were missing were not included (none involved human parainfluenza-positive specimens).

(72%) occurred before April 1, 2020, and the highest HPIV-positive percentage was observed in family shelters (Table).

Six of 32 encounters involved viral co-infections with HPIV (rhinovirus, adenovirus, human bocavi-

rus, enterovirus, and human parechovirus). Participants with HPIV infection reported symptoms at 25 (78%) encounters. Commonly reported symptoms included rhinorrhea (95%), cough (74%), sore throat (53%), and subjective fevers (47%) (Appendix Table

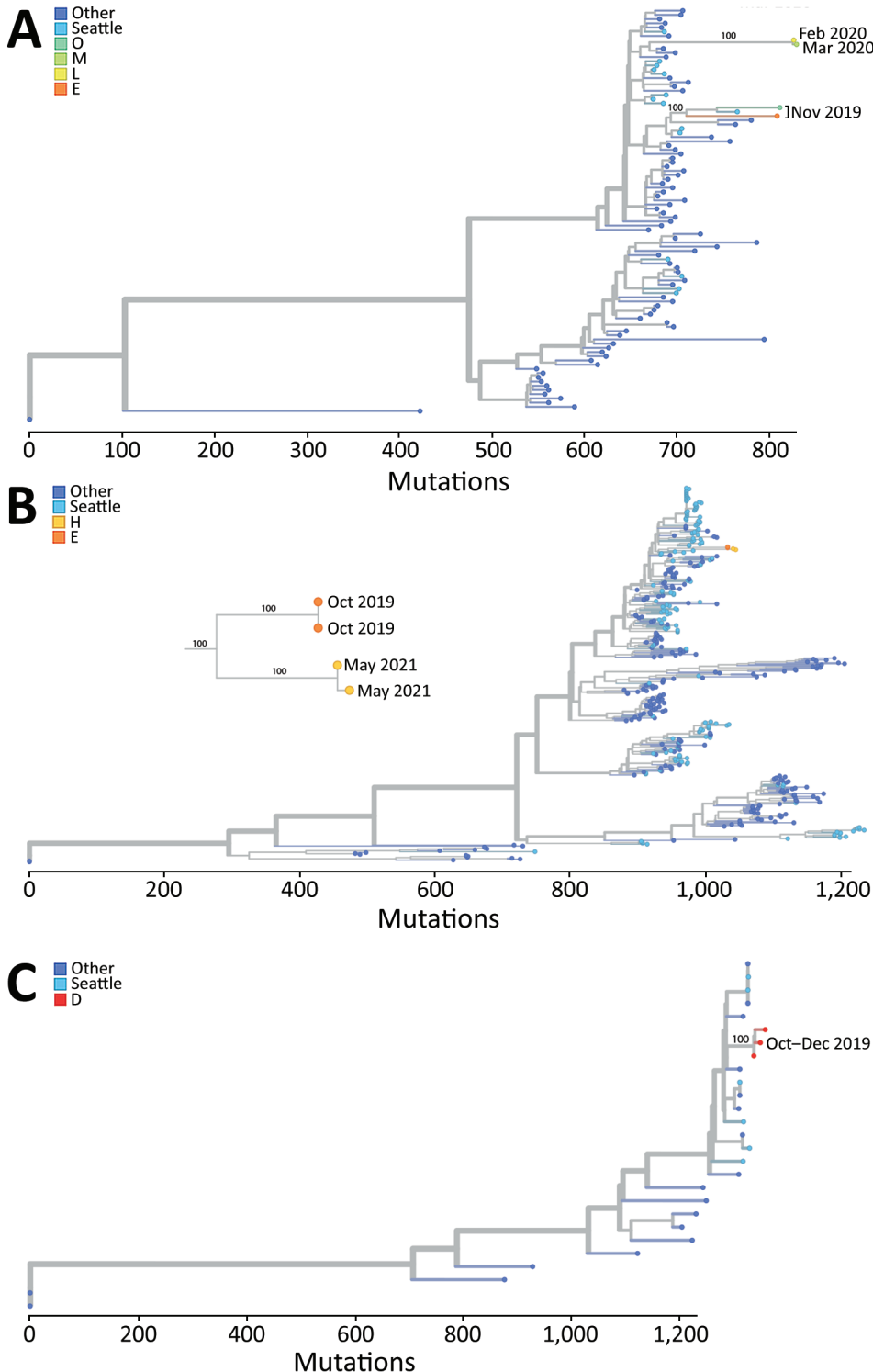


Figure. Phylogenetic trees of human parainfluenza viruses in homeless shelters, King County, Washington, USA, October 2019–May 2021. A) Human parainfluenza virus 1; B) human parainfluenza virus 3; C) human parainfluenza virus 4a. Letters in keys indicate different homeless shelters from which sequenced specimens were collected. Other indicates genomic data from locations not in Seattle, Washington. Seattle indicates genomic data from Seattle other than homeless shelters in this study.

3). HPIV-positive specimens occurred every month during October 2019–April 2020 (Appendix Figure 2). Only 2 HPIV infections were identified during May 2020–April 2021, despite an average of 954 monthly encounters. Six HPIV infections occurred during May 2021 (Appendix Figure 3).

Of 32 HPIV-positive specimens, we identified 3 of the 4 HPIV serotypes: 10 HPIV-1, 11 HPIV-3, and 5 HPIV-4. Six specimens were untypeable. Sequencing of 16 specimens generated 11 sequences (4 HPIV-1, 4 HPIV-3, and 3 HPIV-4a) from 6 shelters (Figure). HPIV-1 sequences formed 2 clusters (100% bootstrap support for each cluster) by collection date in a maximum-likelihood tree that included 94 GenBank HPIV-1 genomes. Both HPIV-3 and HPIV-4a sequences formed single genetic clusters (100% bootstrap support for each cluster) in a maximum-likelihood tree that included 397 GenBank HPIV-3 and 24 HPIV-4a genomes. The HPIV-3 clusters involved HPIV-positive specimens from shelters E (October 2019) and H (May 2021); both shelters housed adults and children. In shelter E, HPIV-3-positive specimens resulted from 6 encounters involving 5 unique participants (all children) spanning 9 days, and 2 specimens were sequenced. In shelter H, 5 HPIV-3 encounters involving 4 unique participants (all children) spanned 17 days, and 2 specimens were sequenced. The sequenced HPIV-3 specimens from shelters E and H, each from unique persons, formed 2 subclusters, each with 100% bootstrap support, corresponding to shelter and collection date.

Respiratory viruses are increasingly appreciated as major pathogens in homeless shelters (5,6). We identified HPIV infections in shelter residents of all ages, although predominantly in children. Family shelters that have mixed populations of adults and children had the greatest percentage of HPIV detections. Two pediatric HPIV-3 clusters occurred before and during the COVID-19 pandemic with genetic clustering by shelter. After the Washington stay-at-home ordinance on March 23, 2020, overall numbers of HPIV infections decreased. These reductions (7) were probably in part caused by community implementation of NPIs because respiratory droplets are probably the main mode of HPIV transmission (8). However, HPIV has been detected on environmental surfaces (9), and shelter site resources might not enable adequate social distancing and air quality.

The pediatric HPIV-3 cases illustrate the need for mitigation guidance to reduce intrashelter HPIV transmission, particularly because younger children have higher upper respiratory tract viral levels than

older persons (10). Limitations of this study included potential selection bias, a lack of site-specific NPI data, cross-sectional study design, and inability to compare concurrent shelter results to community HPIV epidemiology. These HPIV data provide information on site-specific characteristics to inform public health guidance.

The University of Washington Institutional Review Board (study no. 00007800) approved this study.

This study was supported by Gates Ventures, the Centers for Disease Control and Prevention (research contract no. 75D30120C09322 AM002 to H.Y.C.), and the National Institute of Allergy and Infectious Diseases, National Institutes of Health (grant no. T32 AI007044 to E.J.C.).

E.J.C. reported honoraria from Providence Health and Services, Seattle, Washington, for presentations on COVID-19; S.N.C. reported honoraria from University of California, Berkeley, for presentations on COVID-19; A.L.G. reported contract testing for Abbott, Cepheid, Novavax, Pfizer, Janssen, and Hologic and research support from Gilead and Merck, outside of the submitted work; P.R. reported honoraria from The Bill and Melinda Gates Foundation for presentations on COVID-19; J.A.E. reported consulting with Sanofi Pasteur, AstraZeneca, and Meissa Vaccines, and has received research funding from AstraZeneca, GlaxoSmithKline, Merck, and Pfizer outside the submitted work; and H.Y.C. reported consulting with Ellume, Pfizer, The Bill and Melinda Gates Foundation, Glaxo Smith Kline, and Merck, has received research funding from Gates Ventures and Sanofi Pasteur, and support and reagents from Ellume and Cepheid outside of the submitted work.

About the Author

Dr. Chow is the chief of communicable disease epidemiology and immunizations at Public Health Seattle and King County, Seattle, WA. His primary research interests include respiratory virus epidemiology in community settings.

References

1. Wang X, Li Y, Deloria-Knoll M, Madhi SA, Cohen C, Arguelles VL, et al.; Respiratory Virus Global Epidemiology Network. Global burden of acute lower respiratory infection associated with human parainfluenza virus in children younger than 5 years for 2018: a systematic review and meta-analysis. *Lancet Glob Health*. 2021;9:e1077–87. [https://doi.org/10.1016/S2214-109X\(21\)00218-7](https://doi.org/10.1016/S2214-109X(21)00218-7)
2. Howard LM, Edwards KM, Zhu Y, Williams DJ, Self WH, Jain S, et al. Parainfluenza virus types 1-3 infections among children and adults hospitalized with community-acquired pneumonia. *Clin Infect Dis*. 2021;73:e4433–43. <https://doi.org/10.1093/cid/ciaa973>

3. Newman KL, Rogers JH, McCulloch D, Wilcox N, Englund JA, Boeckh M, et al.; Seattle Flu Study Investigators. Point-of-care molecular testing and antiviral treatment of influenza in residents of homeless shelters in Seattle, WA: study protocol for a stepped-wedge cluster-randomized controlled trial. *Trials*. 2020;21:956. <https://doi.org/10.1186/s13063-020-04871-5>
4. Rogers JH, Link AC, McCulloch D, Brandstetter E, Newman KL, Jackson ML, et al.; Seattle Flu Study Investigators. Characteristics of COVID-19 in homeless shelters: a community-based surveillance study. *Ann Intern Med*. 2021;174:42–9. <https://doi.org/10.7326/M20-3799>
5. Chow EJ, Casto AM, Roychoudhury P, Han PD, Xie H, Pfau B, et al. The clinical and genomic epidemiology of rhinovirus in homeless shelters, King County, Washington. *J Infect Dis*. 2022;jiac239. <https://doi.org/10.1093/infdis/jiac239>
6. Chow EJ, Casto AM, Rogers JH, Roychoudhury P, Han PD, Xie H, et al. The clinical and genomic epidemiology of seasonal human coronaviruses in congregate homeless shelter settings: A repeated cross-sectional study. *Lancet Reg Health Am*. 2022;15:100348. <https://doi.org/10.1016/j.lana.2022.100348>
7. Olsen SJ, Winn AK, Budd AP, Prill MM, Steel J, Midgley CM, et al. Changes in influenza and other respiratory virus activity during the COVID-19 pandemic-United States, 2020–2021. *Am J Transplant*. 2021;21:3481–6. <https://doi.org/10.1111/ajt.16049>
8. Henrickson KJ. Parainfluenza viruses. *Clin Microbiol Rev*. 2003; 16:242–64. <https://doi.org/10.1128/CMR.16.2.242-264.2003>
9. Brady MT, Evans J, Cuartas J. Survival and disinfection of parainfluenza viruses on environmental surfaces. *Am J Infect Control*. 1990;18:18–23. [https://doi.org/10.1016/0196-6553\(90\)90206-8](https://doi.org/10.1016/0196-6553(90)90206-8)
10. Hall CB, Geiman JM, Breese BB, Douglas RG Jr. Parainfluenza viral infections in children: correlation of shedding with clinical manifestations. *J Pediatr*. 1977;91:194–8. [https://doi.org/10.1016/S0022-3476\(77\)80811-1](https://doi.org/10.1016/S0022-3476(77)80811-1)

Address for correspondence: Eric J. Chow, Division of Allergy and Infectious Diseases, University of Washington, 1959 NE Pacific St, Box 356423, Seattle, WA 98195, USA; email: ejchow@uw.edu

Presence of *Spirometra mansonii*, Causative Agent of Sparganosis, in South America

Jan Brabec,¹ Manuel Uribe,¹
Jenny J. Chaparro-Gutiérrez, Carlos Hermosilla

Author affiliations: Biology Centre of the Czech Academy of Sciences, České Budějovice, Czech Republic (J. Brabec); CIBAV Research Group, Universidad de Antioquia, Medellín, Colombia (M. Uribe, J.J. Chaparro-Gutiérrez); Justus Liebig University Giessen, Giessen, Germany (M. Uribe, C. Hermosilla)

DOI: <http://doi.org/10.3201/eid2811.220529>

We report molecular identification of an adult *Spirometra mansonii* tapeworm retrieved from a crab-eating fox (*Cerdocyon thous*) in Colombia, confirming presence of this parasite in South America. This tapeworm is the causative agent of human sparganosis, commonly reported from Southeast Asia, and represents the second congeneric species with known zoonotic potential in the Americas.

Sparganosis is a neglected human zoonosis caused by migrating larval stages of the broad tapeworm genus *Spirometra* (Diphyllobothriidea), whose natural definitive hosts include wild and domestic canids and felids. The life cycle of this tapeworm involves 2 intermediate hosts: a freshwater copepod crustacean as the first and various vertebrates, mostly amphibians, as the second. Human infections are commonly reported from Southeast Asia and propagate most often in the form of subcutaneous sparganosis; however, the larvae can enter other organs or parts of central nervous system and cause damage.

Taxonomy of *Spirometra* remains highly complicated. Numerous species of *Spirometra* have been described, often poorly (1), and representatives of just 6 species-level lineages have been characterized molecularly so far, a key prerequisite to achieve a convincing tapeworm identification when only strobila fragments or larval stages are available. Limitations of morphologic characters of *Spirometra* are numerous and include characters' great intraspecific and even intra-individual variability (overview of problematic traits in 2). Molecular sequence data thus represent the only unequivocal method of species identification.

Previous phylogenetic analysis of *Spirometra* has shown that the geographic distribution of the 6 lineages respects continental borders (2). North

¹These authors contributed equally to this article.