SARS-CoV-2 Sequence Analysis during COVID-19 Case Surge, Liberia, 2021

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In June 2021, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) cases surged in Liberia. SARS-CoV-2 sequences from patients hospitalized during March–July 2021 revealed the Delta variant was in Liberia in early March and was dominant in June, irrespective of geography. Mutations and deletions suggest multiple SARS-CoV-2 Delta variant introductions.

Before May 2021, Liberia reported <10 coronavirus disease (COVID-19) cases per day among its population of \approx 5 million (1). Thereafter, case numbers, hospitalizations, and deaths rapidly increased and peaked to >200 cases and 10–15 deaths per day in mid-July 2021 (Appendix Figure 1, https://wwwnc.cdc.gov/EID/article/27/12/21-1818-App1.pdf). To determine whether the rapid case surge was associated with the introduction of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variants of concern or newly emerging variants, we collected nasopharyngeal swab samples from 267 hospitalized patients countrywide during March-July 2021 for high-throughput sequencing.

We collected samples in viral transport media from Bomi, Bong, Grand Cape Mount, Lofa, Margibi, Maryland, Montserrado, and Nimba Counties (Appendix Figure 2). We noted sample collection date and site and sex and median age of patients from whom samples were obtained (Table; Appendix Table). We used Buffer AVL (QIAGEN, https:// www.qiagen.com) lysis buffer to extract total nucleic acid and performed PCR by using the Triplex-CII-SARS-Cov-2 rRT PCR assay (2). We conducted further high-throughput sequencing on 89/267 (33.3%) samples that had cycle threshold values <33 (Appendix Table).

To prepare libraries, we used the Kapa Hyperplus Kit (Roche, https://www.roche.com) on first strand cDNA synthesized from 89 RNA samples (3), then we enriched for SARS-CoV-2 by using myBaits Custom RNA-Seq Kit (Daicel Arbor Biosciences, https://arborbiosci.com). We sequenced captured libraries on Nextseq 2000 or Nextseq 550 (Illumina, https://www.illumina.com), which yielded 5–8 million 220-bp reads per sample. We mapped reads to a SARS-CoV-2 reference sequence (GenBank accession no. NC_045512) to determine variants (Table; Appendix Table).

Of the 89 RNA samples, 77 (86.5%) yielded complete coding sequences with a minimum depth of $\approx 15 \times$ (GISAID accession nos. EPI_ISL_3547663-705, EPI_ISL_3560291, and EPI_ISL_4232122-52). Using high-throughput sequencing data, we generated consensus fasta sequences of 77 SARS-CoV-2 genomic sequences and further analyzed sequences by using Geneious R10 (https://www.geneious.com), Next-Strain (4), and GISAID (5).

Among 77 genomes recovered, 4 (5.2%) were Alpha variant (B.1.1.7); 6 (7.8%) were Beta variant (B.1.351); 1 (1.3%) was Iota variant (B.1.526); 6 (7.8%) were Eta variant (B.1.525); and 56 (72.7%) were Delta variant (B.1.617.2) viruses (Table). We identified Delta variant viruses in samples collected in early March and in April and May 2021, from Bong County. Delta variant viruses were co-circulating with Alpha, Beta, Eta, Iota, and other 20B variant viruses in Liberia. All 44 sequences recovered during June-July 2021 were from Delta variant viruses (Table). We used complete polyprotein coding sequences from Liberia, other representative SARS-CoV-2 sequences, and variant reference sequences to create a maximum-likelihood, nucleotide-based phylogenetic tree in MEGA X (6) (Figure).

Using reference sequence NC_045512 as a baseline, we found 3 Alpha variant-specific amino acid deletions (H69del, V70del, Y144del) in the surface glycoprotein of all Alpha variant genomes and 3 Beta variant-specific amino acid deletions (L241del, L242del, A243del) in the surface glycoprotein of all Beta variant genomes. All 56 Delta variant genomes had the 2 variant-specific amino acid deletions, F157del and R158del, and 8 of 9 other Delta variant-

¹These first authors contributed equally to this article.

couling genomic sequences, Liberta, 2021											
					No.	SARS-CoV-2 variant, no. of samples/county/mo					
Month	Total no.	Patient	Average age,		samples/	Delta	Alpha	Beta	Eta	lota	20B
collected	samples	sex, no.	y (SD)	County	county	B.1.617.2	B.1.1.7	B.1.351	B.1.525	B.1.526	other
Mar	4	2M, 2F	39.25 (6.05)	Montserrado	3			1	1	1	
			. ,	Bong	1	1					
Apr	11	10M, 1F	42.54 (11.52)	Montserrado	10	4	1	3	2		-
				Grand Cape	1			1			
				Mount							
May	18	9M, 9F	40.11 (16.82)	Bong	1	1					
-				Margibi	1						
				Montserrado	14	6	3	2	2		1
				Nimba	2						2
Jun	36	13M, 23F	39.22 (18.36)	Lofa	5	5					-
				Margibi	1	1					
				Maryland	1	1					
				Montserrado	29	29					
Jul	8	4M, 4F	51.25 (9.71)	Margibi	1	1					
				Montserrado	5	5					
				Nimba	2	2					
*Liberia experienced a surge in COVID-19 cases during June 2021. Blank cells indicate no variants detected. COVID-19, coronavirus disease; SARS-											
CoV-2, severe acute respiratory syndrome coronavirus 2.											

Table. Characteristics of 77 clinical samples collected before and during COVID-19 case surge that yielded complete SARS-CoV-2 coding genomic sequences, Liberia, 2021

specific amino acid substitutions in the surface glycoprotein (T19R, G142D, E156G, L452R, T478K, D614G, P681R, and D950N). The A222V surface glycoprotein mutation was absent in only 2/56 Delta variant genomes, LIB-0226 and LIB-0217, collected from Monteserrado County in May 2021 (4). We observed another mutation in the surface glycoprotein, V367L, in 14 sequences: 1 from Bong, 2 from Margibi, 1 from Maryland, 9 from Montserrado, and 1 from Nimba. No sequences recovered from Lofa County had the V367L mutation. We noted the R724K mutation in the open reading frame 1a region of 2 sequences from Lofa, LIB-0131 and LIB-0133. LIB-0073 and LIB-0093 sequences collected from Montserrado County had 2 amino acid deletions in the open reading frame 8 region (position 120–121).

Recent surges in COVID-19 in many countries have been associated with the emergence of highly transmissible Delta variant viruses (7,8). In March 2021, the National Public Health Institute of Liberia sequenced 10 random samples from hospitalized COVID-19 patients in Monteserrado; all sequences were Alpha variant viruses (B. Shobayo, unpub. data).

A limitation of our study is the small sample sets used for analysis; nonetheless, our findings suggest that Alpha and other circulating variant viruses were replaced by Delta variant viruses countrywide in Liberia in <3 months. Mutation and phylogenetic analyses further indicate that several Delta variant strains were circulating after March 2021 and suggest multiple separate introductions.

Before June 2021, only a small percentage of the population was vaccinated in Liberia. The infections we report occurred in unvaccinated persons. The Ministry of Health, Liberia, initiated a vaccination drive in August 2021. By September, ≈130,000 persons, >2% of the population, had received a single dose of the Johnson & Johnson/Janssen vaccine (https://www.jnj.com). The COVID-19 vaccination campaign is ramping up as <30 cases/day are reported in Liberia, but the currently circulating Delta variants are a concern because they contain mutations and deletions in the surface glycoprotein that might influence vaccine efficacy (9). Liberia should continued surveillance for SARS-CoV-2 variants of concern to determine whether additional vaccination or public health measures are needed to curb severe disease and future case surges in the country.

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Dr. Shobayo is public health and medical research scientist and deputy director at National Public Health Institute of Liberia, Monrovia, Liberia, and member of Partnership for Research on Infectious Diseases in Liberia. His research interests include survillence and investigation of emerging and re-emerging viruses and their impacts on public health.



Figure. Phylogenetic analysis of 77 nasopharyngeal swab samples collected during coronavirus disease case surge, Liberia, March– July 2021, and reference sequences. We created a maximum-likelihood nucleotide phylogenetic tree of the complete polyprotein coding region by using MEGA X (https://www.megasoftware.net), with a bootstrap value of 100 and and used Tamura-Nei 93 (TN93) as a substitution model with a discrete gamma distribution (+G) for evolutionary rate; the rate variation model allowed some sites to be evolutionarily invariable (+I). Numbers along the branches are bootstrap values of 100 bootstrap resamplings. Teal indicates samples collected in March 2021; purple indicates samples collected in April 2021; pink indicates samples collected in May 2021; blue indicates samples collected in June 2021; orange indicates samples collected in July 2021; brown indicates variants of concern or variants of interest; black indicates other circulating variants; green indicates severe acute respiratory syndrome coronavirus 2 reference sequence and other early parental sequences from 2020.

References:

- Worldometer. COVID-19 coronavirs disease data, Liberia [cited 2021 Sep 26]. https://www.worldometers.info/ coronavirus/country/liberia
- 2. US Food and Drug Administration. Accelerated emergency use authorization (EUA) summary the Triplex CII-SARS-CoV-2 rRT-PCR test updated 9/14/2020 [cited 2021 Sep 15]. https://www.fda.gov/media/137983/download
- 3. Mishra N, Ng TFF, Marine RL, Jain K, Ng J, Thakkar R, et al. Antibodies to enteroviruses in cerebrospinal fluid of patients with acute flaccid myelitis. MBio. 2019;10:e01903-19. https://doi.org/10.1128/mBio.01903-19
- GIŜAID. Tracking of variants [cited 2021 Sep 15]. https://www.gisaid.org/hcov19-variants
- 5. Next-Strain. Real-time tracking of pathogen evolution [cited 2021 Sep 15]. https://nextstrain.org
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. Mol Biol Evol. 2018;35:1547–9. https://doi.org/ 10.1093/molbev/msy096
- Lopez Bernal J, Andrews N, Gower C, Gallagher E, Simmons R, Thelwall S, et al. Effectiveness of Covid-19 vaccines against the B.1.617.2 (Delta) variant. N Engl J Med. 2021;385:585–94. https://doi.org/10.1056/NEJMoa2108891
- Alizon S, Haim-Boukobza S, Foulongne V, Verdurme L, Trombert-Paolantoni S, Lecorche E, et al. Rapid spread of the SARS-CoV-2 Delta variant in some French regions, June 2021. Euro Surveill. 2021;26.2100573. https://doi.org/ 10.2807/1560-7917.ES.2021.26.28.2100573
- Creech CB, Walker SC, Samuels RJ. SARS-CoV-2 vaccines. JAMA. 2021;325:1318–20. https://doi.org/10.1001/ jama.2021.3199

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Real-Time Projections of SARS-CoV-2 B.1.1.7 Variant in a University Setting, Texas, USA

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We used the incidence of spike gene target failures identified during PCR testing to provide an early projection of the prevalence of severe acute respiratory syndrome coronavirus 2 variant B.1.1.7 in a university setting in Texas, USA, before sequencing results were available. Findings from a more recent evaluation validated those early projections.

dentification of the highly transmissible novel se-Lvere acute respiratory syndrome coronavirus 2 (SARS-CoV-2) variant B.1.1.7 (Alpha variant) in the United Kingdom raised concerns for renewed pandemic surges worldwide (1,2). B.1.1.7 likely arrived in the United States by October 2020 (1); it was first detected in December 2020 and declared the dominant strain in April 2021, as projected in January 2021 (3). However, the regional prevalence of B.1.1.7 was largely unknown in early 2021 because of limited molecular surveillance for SARS-CoV-2 (4). To provide local situational awareness at that pivotal moment in the coronavirus disease (COVID-19) pandemic, we estimated the prevalence of B.1.1.7 on the basis of 17,003 student SARS-CoV-2 PCR test results reported through the Proactive Community Testing Program at the University of Texas (UT; Austin, Texas, USA), a large public university located in a metropolitan area with a population >2 million, during January 16-February 12, 2021 (K.E. Johnson et al., unpub. data, https://doi.org/10.1101/2021.03.05.21252541. Those early estimates were subsequently validated by using PCR data through April 9, 2021.

Mutations in the B.1.1.7 spike protein result in a failure to detect the spike gene probe in standard SARS-CoV-2 quantitative reverse transcription PCR (qRT-PCR). In estimating the prevalence of B.1.1.7 from local quantitative PCR data, we initially assumed US estimates for the proportion of spike gene target failures (SGTF) attributable to B.1.1.7 (4) and, in our retrospective analysis, update that proportion on the basis of local sequencing data. We used a Bayesian model to estimate the local growth rate of B.1.1.7 among all SARS-CoV-2 infections and applied a compartmental susceptible-exposed-infected-recovered model of SARS-CoV-2 transmission to project the effect of B.1.1.7 on future COVID-19 prevalence.

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Appendix

Appendix Table. Dates, collection sites, sex, age, quantitative PCR cycle threshold values, and sequence coverage for 89 samples from COVID-19 patients, Liberia, 2021

	Collection	*	Age,		% Genome	Avg.			GISAID
Sample ID	date	County	y/sex	C _t value	recovered	depth/nucleotide	Nextclade	Variant†	clade
LIB-0233	9 Mar	Bong	47/M	32.39	99.82	53.35	21A	VOC Delta B.1.617.2	G
LIB-0193	10 Mar	Montserrado	30/F	32.70	99.73	29.48	21D	VOI Eta B.1.525	G
LIB-0166	31 Mar	Montserrado	40/F	26.94	99.94	1534.63	20H	VOC Beta B 1 351	GH
LIB-0200	31 Mar	Montserrado	40/M	23.43	99.94	14633.94	21F	VOI lota B 1 526	GH
LIB-0170	2 Apr	Montserrado	58/M	24.77	99.97	24611.29	20H	VOC Beta B 1 351	GH
LIB-0171	4 Apr	Montserrado	64/F	25.04	99.86	185.61	21D	VOI Eta B 1 525	G
LIB-0174	4 Apr	Montserrado	35/M	24.63	99.98	27842.07	201	VOC Alpha B 1 1 7	GR
LIB-0192	4 Apr	Montserrado	30/M	22.07	100.00	69811.5	20H	VOC Beta B 1 351	GH
LIB-0197	6 Apr	Montserrado	54/M	23.87	99.96	16087.41	21A	VOC Delta	G
LIB-0152	8 Apr	Montserrado	41/M	24 83	99 91	681 27	20B	Other	GR
LIB-0173	8 Apr	Grand Cape Mount	41/M	27.23	99.94	2708	20H	VOC Beta B 1 351	GH
LIB-0196	10 Apr	Montserrado	30/M	19.29	100.00	93864.59	21A	VOC Delta B.1.617.2	G
LIB-0198	10 Apr	Montserrado	27/M	24.05	99.98	11960.47	21A	VOC Delta B.1.617.2	G
LIB-0201	13 Apr	Montserrado	46/M	26.42	99.94	2851.87	21D	VOI Eta B.1.525	G
LIB-0182	17 Apr	Montserrado	42/M	23.72	99.99	22901.55	21A	VOC Delta B.1.617.2	G
LIB-0204	3 May	Montserrado	20/M	32.16	43.18	1.62	NA	NA	NA
LIB-0220	3 May	Montserrado	34/M	29.37	99.92	215.57	201	VOC Alpha B.1.1.7	GR
LIB-0248	4 May	Montserrado	30/M	31.31	99.85	100.86	21A	VOC Delta B.1.617.2	G
LIB-0206	15 May	Montserrado	46/M	28.51	57.74	1.99	NA	NA	NA
LIB-0162	17 May	Margibi	77/M	32.11	99.85	434.73	21D	VOI Eta B.1.525	G
LIB-0236	17 May	Nimba	62/F	23.46	99.94	4894.08	20B	Other	GR
LIB-0203	20 May	Nimba	34/M	32.15	99.71	39.59	20B	Other	GR
LIB-0205	20 May	Montserrado	64/F	31.14	99.88	64.74	201	VOC Alpha B.1.1.7	GR
LIB-0217	20 May	Montserrado	37/M	30.01	99.89	175.39	21A	VOC Delta B.1.617.2	G
LIB-0218	20 May	Montserrado	24/F	32.81	99.81	18.65	21A	VOC Delta B.1.617.2	G
LIB-0221	20 May	Montserrado	10/F	31.91	99.71	14.87	201	VOC Alpha B.1.1.7	GR
LIB-0225	20 May	Montserrado	68/M	32.97	85.36	2.78	NA	NA	NA
LIB-0226	20 May	Montserrado	63/M	32.64	99.80	34.1	21A	VOC Delta B.1.617.2	G

	Collection		Aae.		% Genome	Ava.			GISAID
Sample ID	date	County	y/sex	C _t value	recovered	depth/nucleotide	Nextclade	Variant†	clade
LIB-0247	20 May	Montserrado	43/F	18.86	100.00	303638.25	20H	VOC Beta	GH
			00/F	00 7 0	~~~~	44040.00	005	B.1.351	0.5
LIB-0250	20 May 20 May	Montserrado	38/F 33/F	22.72	99.98	14212.06	20B 21A	Uther	GR
LID-0251	20 May	Monisenado	33/F	20.09	100.00	7020.71	ZIA	B 1 617 2	G
LIB-0255	20 May	Montserrado	31/M	17.25	100.00	504226.47	20H	VOC Beta	GH
								B.1.351	
LIB-0253	27 May	Montserrado	27/F	18.58	100.00	223902.08	21D	VOI Eta	G
	20 May	Pong	54/NA	16.01	100.00	102424 00	21 4	B.1.525	C
LID-0240	29 Way	вону	54/10	10.01	100.00	193424.09	21A	B 1 617 2	G
LIB-0242	29 May	Montserrado	23/F	22.93	99.99	18571.69	21D	VOI Eta	G
								B.1.525	
LIB-0244	29 May	Montserrado	38/M	19.90	100.00	15944.35	21A	VOC Delta	G
	5 lup	Montsorrado	57/M	21	100.00	122105 27	21.0	B.1.617.2	G
LID-0004	5 5011	Monisenado	57/101	21	100.00	132103.37	214	B 1 617 2	9
LIB-0007	5 Jun	Montserrado	67/F	21	100.00	24117.24	21A	VOC Delta	G
								B.1.617.2	
LIB-0071	5 Jun	Montserrado	28/M	28	99.98	748.95	21A	VOC Delta	G
	6 lup	Montsorrado	62/E	28	00.04	302 55	21 A	B.1.617.2	G
	0 Juli	Monisenado	02/1	20	33.34	302.33	214	B.1.617.2	9
LIB-0018	6 Jun	Montserrado	41/M	24	99.30	106.19	21A	VOC Delta	G
								B.1.617.2	
LIB-0019	6 Jun	Montserrado	34/M	17	99.90	359124.54	21A	VOC Delta	G
LIB-0021	6 lun	Montserrado	56/F	22	100.00	24466 47	21Δ	VOC Delta	G
LID-0021	0 0011	Montschado	50/1	22	100.00	24400.47	217	B.1.617.2	0
LIB-0022	6 Jun	Montserrado	37/F	25	100.00	1307.05	21A	VOC Delta	G
=								B.1.617.2	-
LIB-0026	6 Jun	Montserrado	38/M	19	99.90	366820.93	21A	VOC Delta	G
LIB-0028	6.lun	Montserrado	27/F	27	99 94	797 2	21A	VOC Delta	G
212 0020	0 0 dill	montoonado	2.7.		00.01	101.2	200	B.1.617.2	U
LIB-0032	6 Jun	Margibi	46/F	23	100.00	30961.5	21A	VOC Delta	G
	0.1		40/5	00	00.05	000.0	014	B.1.617.2	0
LIB-0064	8 Jun	Montserrado	40/F	28	99.95	399.9	21A	VOC Delta	G
LIB-0012	12 Jun	Montserrado	45/F	19	100.00	12751.38	21A	VOC Delta	G
						12101100		B.1.617.2	U U
LIB-0029	12 Jun	Montserrado	30/M	23	100.00	6319.4	21A	VOC Delta	G
	10 1	Manualanad	00/5	05	100.00	7005 40	014	B.1.617.2	0
LIB-0030	12 Jun	Maryland	32/F	25	100.00	7395.49	21A	R 1 617 2	G
LIB-0037	12 Jun	Montserrado	35/M	20	100.00	2987.68	21A	VOC Delta	G
								B.1.617.2	
LIB-0040	12 Jun	Montserrado	38/M	22	100.00	40117.32	21A	VOC Delta	G
	12 lun	Montsorrado	20/M	22	100.00	22265 27	21 A	B.1.617.2	G
LID-0042	12 Juli	Monisenado	29/101	22	100.00	23303.37	214	B 1 617 2	9
LIB-0045	12 Jun	Montserrado	32/F	20	100.00	10116.23	21A	VOC Delta	G
								B.1.617.2	
LIB-0047	12 Jun	Montserrado	32/F	22	100.00	46601.79	21A	VOC Delta	G
LIB-0128	15 Jun	Lofa	1/M	21	100.00	29163 85	214	VOC Delta	G
LID-0120	10 0011	Lola	1/101	21	100.00	20100.00	217	B.1.617.2	0
LIB-0010	16 Jun	Montserrado	26/F	24	100.00	9787.07	21A	VOC Delta	G
			F0/-	05	400.00	1001.11	644	B.1.617.2	~
LIB-0131	16 Jun	Lota	56/F	25	100.00	1864.44	21A	VOC Delta	G
LIB-0133	16 Jun	Lofa	47/F	26	99,99	65698 71	21A	VOC Delta	G
				20				B.1.617.2	-
LIB-0136	16 Jun	Lofa	35/M	22	99.98	1442.69	21A	VOC Delta	G
	16	Dam!	40/5	05	0	NIA	NIA	B.1.617.2	NIA
LID-0137	16 Jun	l ofa	40/F 30/F	∠5 23	0 99.95	INA 5513	1NA 21A	INA VOC Delta	INA G
				20				B.1.617.2	-

	Collection		Age		% Genome	Ανα			GISAID
Sample ID	date	County	v/sex	C₊ value	recovered	depth/nucleotide	Nextclade	Variant†	clade
LIB-0118	17 Jun	Montserrado	41/M	19	100.00	74762.42	21A	VOC Delta	G
								B.1.617.2	
LIB-0121	17 Jun	Montserrado	82/F	30	99.66	40	21A	VOC Delta	G
								B.1.617.2	
LIB-0123	17 Jun	Montserrado	25/F	21	100.00	10448.21	21A	VOC Delta	G
								B.1.617.2	
LIB-0135	17 Jun	Montserrado	55/F	23	99.97	598.15	21A	VOC Delta	G
								B.1.617.2	-
LIB-0099	18 Jun	Montserrado	2/F	21	100.00	12031.71	21A	VOC Delta	G
	40 1	Manufa anna da	05/5	00	00.00	447.00	014	B.1.617.2	0
LIB-0124	18 Jun	Montserrado	25/F	29	99.88	117.63	21A	VUC Delta	G
	19 Jun	Montoorrado	90/E	22	100.00	12009 5	21 4		C
LID-0125	To Juli	wontsenado	09/F	22	100.00	13006.5	21A	R 1 617 2	G
LIB-0102	19 lun	Montserrado	8/F	20	100.00	10170.05	214	VOC Delta	G
LID-0102	10 0011	Montschado	0/1	20	100.00	10170.00	217	B 1 617 2	0
LIB-0130	24 Jun	Montserrado	37/M	30	71 32	NA	NA	NA	NA
LIB-0134	24 Jun	Montserrado	62/F	28	12.74	NA	NA	NA	NA
LIB-0142	26 Jun	Montserrado	42/F	27	5.53	NA	NA	NA	NA
LIB-0147	26 Jun	Montserrado	36/M	30	99.97	1970.45	21A	VOC Delta	G
								B.1.617.2	
LIB-0069	27 Jun	Montserrado	48/F	19	100.00	86159.99	21A	VOC Delta	G
								B.1.617.2	
LIB-0111	7 Jul	Montserrado	11/F	29	0.65	0.9	NA	NA	NA
LIB-0113	7 Jul	Montserrado	48/M	26	1.65	0.86	NA	NA	NA
LIB-0073	8 Jul	Montserrado	31/F	29	99.47	117.79	21A	VOC Delta	G
								B.1.617.2	-
LIB-0074	8 Jul	Montserrado	47/M	24	100.00	4168.52	21A	VOC Delta	G
	0.1.1	Manufa anna da	F 4 / F	00	400.00	1 1000 00	014	B.1.617.2	0
LIB-0076	8 Jui	Montserrado	51/F	22	100.00	14232.36	21A	VUC Delta	G
	0 1.1	Moraihi	54/E	22	100.00	12001 24	21 4		C
LID-0070	o Jui	wargibi	04/F	23	100.00	12901.34	21A	R 1 617 2	G
1 IB-0093	8 Jul	Montserrado	48/M	27	00 31	84 44	214	VOC Delta	G
LID-0000	0.001	Montschado	40/10	21	55.51	0-1-1-	217	B 1 617 2	0
LIB-0103	8 Jul	Montserrado	41/F	29	0	NA	NA	NA	NA
LIB-0112	8 Jul	Montserrado	32/F	29	0 67	1 67	NA	NA	NA
LIB-0084	9 Jul	Nimba	67/M	27	99.91	1228.28	21A	VOC Delta	G
								B.1.617.2	
LIB-0085	9 Jul	Nimba	59/M	22	100.00	63134.43	21A	VOC Delta	G
								B.1.617.2	
LIB-0094	10 Jul	Montserrado	52/F	32	99.93	1979.92	21A	VOC Delta	G
								B.1.617.2	
LIB-0101	20 Jul	Montserrado	64/M	31	26.44	NA	NA	NA	NA

*Twelve samples had insufficient genome coverage of average 15× depth/nucleotide.Ct, cycle threshold; NA, not applicable; VOC, variant of concern; VOI, variant of interest. Sequencing data can be accessed using GISAID (https://www.gisaid.org) accession numbers EPI_ISL_3547663–705, EPI_ISL_3560291, and EPI_ISL_4232122–52). †According to Phylogenetic Assignment of Named Global Outbreak Lineages (https://cov-lineages.org) software tool.



Appendix Figure 1. Daily coronavirus disease cases, Liberia, February 2020–September 2021. Blue lines indicate 7 day moving average of daily new cases. Data were derived from Worldometer (https://www.worldometers.info/coronavirus/country/liberia).



Appendix Figure 2. Sites from which nasopharyngeal swab samples were collected during a coronavirus disease case surge, Liberia, 2021. Red stars indicate locations inside counties; black star indicates the capital, Monrovia. Numbers of cases per county are indicated in parentheses.