

under programmatic conditions to improve understanding of the recommended treatment regimens, their implementation, and the effects on TB management globally.

This research was supported by the grants from the National Institute of Allergy and Infectious Diseases/US Civilian Research & Development Foundation, CRDF Global (DAA9-19-66199) and the Ministry of Health of Ukraine (#0123100178). The study funders had no role in the study design, data collection, data analysis, data interpretation, or writing of the report.

### About the Author

Dr. Dahl is a medical doctor and PhD student at the Department of Infectious Diseases, Aarhus University Hospital, Aarhus, Denmark. He is interested in complicated respiratory infections such as tuberculosis and nontuberculous mycobacteria and mainly works with clinical epidemiology and systematic reviews.

### References

1. World Health Organization. Meeting report of the WHO expert consultation on the definition of extensively drug-resistant tuberculosis [cited 2023 Dec 6]. <https://www.who.int/publications/i/item/9789240018662>
2. Roelens M, Battista Migliori G, Rozanova L, Estill J, Campbell JR, Cegielski JP, et al. Evidence-based definition for extensively drug-resistant tuberculosis. *Am J Respir Crit Care Med*. 2021;204:713–22. <https://doi.org/10.1164/rccm.202009-3527OC>
3. Pedersen OS, Holmgaard FB, Mikkelsen MKD, Lange C, Sotgiu G, Lillebaek T, et al. Global treatment outcomes of extensively drug-resistant tuberculosis in adults: A systematic review and meta-analysis. *J Infect*. 2023;87:177–89. <https://doi.org/10.1016/j.jinf.2023.06.014>
4. Kherabi Y, Fréchet-Jachym M, Rhioux C, Yazdanpanah Y, Méchai F, Pourcher V, et al.; MDR-TB Management Group. Revised definitions of tuberculosis resistance and treatment outcomes, France, 2006–2019. *Emerg Infect Dis*. 2022;28:1796–804. <https://doi.org/10.3201/eid2809.220458>
5. Mikiashvili L, Kempker RR, Chakhaia T, Bablishvili N, Avaliani Z, Lomtadze N, et al. Impact of prior TB treatment with new/companion drugs on clinical outcomes in patients receiving concomitant bedaquiline and delamanid for MDR/RR-TB. *Clin Infect Dis*. 2023 Nov 14 [Epub ahead of print]. <https://doi.org/10.1093/cid/ciad694>
6. Pedersen OS, Butova T, Kapustnyk V, Miasoiedov V, Kuzhko M, Hryshchuk L, et al. Treatment outcomes and risk factors for an unsuccessful outcome among patients with highly drug-resistant tuberculosis in Ukraine. *Clin Microbiol Infect*. 2024;30:360–7. <https://doi.org/10.1016/j.cmi.2023.12.001>
7. World Health Organization. Definitions and reporting framework for tuberculosis – 2013 revision: updated December 2014 and January 2020 [cited 2023 Dec 6]. <https://www.who.int/publications/i/item/9789241505345>
8. Veziris N, Bonnet I, Morel F, Guglielmetti L, Maitre T, Fournier Le Ray L, et al.; CNR MyRMA; Members of the CNR-MyRMA (French National Reference Center for Mycobacteria). Impact of the revised definition of extensively drug-resistant tuberculosis. *Eur Respir J*. 2021;58:2100641 <https://doi.org/10.1183/13993003.00641-2021>
9. Collaborative Group for the Meta-Analysis of Individual Patient Data in MDR-TB treatment–2017; Ahmad N, Ahuja SD, Akkerman OW, et al. Treatment correlates of successful outcomes in pulmonary multidrug-resistant tuberculosis: an individual patient data meta-analysis. *Lancet*. 2018;392:821–34. [https://doi.org/10.1016/S0140-6736\(18\)31644-](https://doi.org/10.1016/S0140-6736(18)31644-)
10. Ismail NA, Omar SV, Moultrie H, Bhyat Z, Conradie F, Enwerem M, et al. Assessment of epidemiological and genetic characteristics and clinical outcomes of resistance to bedaquiline in patients treated for rifampicin-resistant tuberculosis: a cross-sectional and longitudinal study. *Lancet Infect Dis*. 2022;22:496–506. [https://doi.org/10.1016/S1473-3099\(21\)00470-9](https://doi.org/10.1016/S1473-3099(21)00470-9)

Address for correspondence: Victor Næstholt Dahl, Department of Infectious Diseases, Aarhus University Hospital, Palle Juul-Jensens Blvd 99, Aarhus, Denmark, DK-8200; email: vicmat@rm.dk

## Opportunistic *Elizabethkingia miricola* Infections in Intensive Care Unit, Spain

Eva Soler-Iborte, Mario Rivera-Izquierdo, Carmen Valero-Ubierna

Author affiliations: Hospital Universitario San Cecilio, Granada, Spain (E. Soler-Iborte, M. Rivera-Izquierdo, C. Valero-Ubierna); Instituto biosanitario de Granada, Granada, (M. Rivera-Izquierdo); Ciber de Epidemiología y Salud Pública, Madrid, Spain (M. Rivera-Izquierdo)

DOI: <https://doi.org/10.3201/eid3004.231491>

In 2021, we identified a cluster of *Elizabethkingia miricola* cases in an intensive care unit in Spain. Because *E. miricola* is not considered a special surveillance agent in Spain, whole-genome sequencing was not performed. The bacterial source was not identified. All *Elizabethkingia* species should be listed as special surveillance bacteria.

The *Elizabethkingia* genus is formed by a group of gram-negative, aerobic, and nonfermenting bacteria widely distributed in nature and environments, such as water and hospital taps (1). In 2003, a new a

bacterial species was identified in the condensation water obtained from the Mir space station in 1997 and was assigned as *Chryseobacterium miricola* (2). That new species was later transferred to the *Elizabethkingia* genus and renamed *Elizabethkingia miricola* (3). This species is considered as an uncommon low-pathogenic agent in clinical samples, acting as an opportunistic pathogen, but since 2008, it has become an emerging bacterium of increasing relevance (4). *E. miricola* has not been fully epidemiologically characterized but is considered intrinsically resistant to multiple drugs (5).

We describe a cluster of *E. miricola* in the intensive care unit (ICU) of the Hospital Universitario San Cecilio in Granada, Spain. The index case corresponded to a 66-year-old man hospitalized for COVID-19. The microbiology service identified *E. miricola* isolates from a bronchial aspirate sample by using matrix-assisted laser desorption/ionization

time-of-flight mass spectrometry and informed the ICU of a positive result on March 19, 2021. Two days later, a 70-year-old man hospitalized in the same unit for COVID-19 also tested positive for *E. miricola* in a bronchial aspirate sample sent for a previous diagnosis with tracheobronchitis associated with *Stenotrophomonas maltophilia*.

Throughout 2021, we found 13 more cases of *E. miricola* in the same ICU. Given the identification of the same species, and for clinical and epidemiologic criteria, the outbreak was considered nosocomial. However, although isolates were retained for potential future research, no whole-genome sequencing could be performed because the species is not included as special surveillance agent for the Andalusian Health System because of resource limitations. The reason for admission in 11 (73.3%) patients was COVID-19. Of the 15 case-patients, 11 (73.3%) were men and 4 (26.7%) women, 6 (40.0%) had tracheobronchitis diagnoses, 3 (20.0%) had

**Table.** Characteristics of case-patients in an outbreak opportunistic *Elizabethkingia miricola* infections in an intensive care unit, Spain\*

Age, y/sex	Isolation sample	ICU stay, d	Diagnosis†	Diagnosis date, 2021	Death	Reason for admission	Antimicrobial drug resistance‡
66/M	BAS	40	Isolation	Mar 17	N	COVID-19	Carbapenems, ceftazidime, cefepime, aztreonam
70/M	BAS	63	Isolation	Mar 19	N	COVID-19	Carbapenems, ceftazidime, cefepime, aztreonam
46/F	BAS	12	Isolation	Mar 27	Y	Carcinosis	Carbapenems, ceftazidime, cefepime, aztreonam, aminoglycosides
64/M	BAS	61	Tracheobronchitis	Apr 22	N	COVID-19	Carbapenems, ceftazidime, cefepime, aztreonam, piperacillin/tazobactam
57/F	BAS, catheter	20	Bacteriemia	Apr 27	Y	COVID-19	Carbapenems, ceftazidime, cefepime, aztreonam, piperacillin/tazobactam, aminoglycosides, trimethoprim/sulfamethoxazole
50/M	BAS	58	Tracheobronchitis	May 19	Y	COVID-19	Carbapenems, ceftazidime, amikacin
52/M	BAS	37	Tracheobronchitis	May 20	N	COVID-19	Carbapenems, ceftazidime, amikacin
56/M	BAS	43	Ventilator-associated pneumonia	May 22	Y	COVID-19	Amikacin
58/F	BAS	80	Tracheobronchitis	May 29	N	COVID-19	Amikacin
58/M	BAS	30	Isolation	Jul 7	Y	Fever	Piperacillin/tazobactam, linezolid
37/F	BAS	51	Isolation	Aug 20	Y	COVID-19	Piperacillin/tazobactam, linezolid
32/M	BAS	93	Tracheobronchitis	Sep 18	Y	COVID-19	Piperacillin/tazobactam, linezolid
74/M	BAS	27	Tracheobronchitis	Sep 28	Y	Epileptic seizures	Piperacillin/tazobactam, linezolid
61/M	BAS	27	Ventilator-associated pneumonia	Oct 27	N	Septic shock	Piperacillin/tazobactam, linezolid, vancomycin,
73/M	BAS	15	Ventilator-associated pneumonia	Dec 30	N	COVID-19	Piperacillin/tazobactam, linezolid, levofloxacin

\*BAS, bronchoaspiration; ICU, intensive care unit.

†Isolation means that no other clinical pathology was reported by the responsible physician in the ICU. Therefore, *E. miricola* identification was considered as asymptomatic colonization.

‡Antibiograms were compatible with the same agent, but development of new resistances was identified during the 9-mo outbreak.

Trimethoprim/sulfamethoxazole showed the best antibiogram sensitivity: 10 patients showed sensitivity, 4 intermediate sensitivity, and only 1 showed resistance. Sensitivity to levofloxacin was observed for 7 (46.7%) patients; 4 more showed intermediate sensitivity.

ventilator-associated pneumonia, and 1 (6.7%) had catheter-related bacteremia; 8 (53.3%) patients died (Table).

All patients received steroid treatment during their ICU stays. All case-patients were intubated during their hospitalization. The average length of ICU stay was 43.8 days, and the length between admission and identification of the agent was long, a mean of 26.4 days.

Because 290 days elapsed from identification of the index case (March 19) to identification of the last case (December 30), we assumed persistence of the agent in the ICU environment. Nevertheless, despite a search of environmental and surface samples, the definitive focus of persistence was not identified. Because of possible cross-transmission in a unit with such vulnerable patients, we notified the Service of Preventive Medicine and Public Health, which initiated prevention measures. Because of the lack of available knowledge related to *E. miricola* and closely related species, we reinforced standard precautions and established contact precautions. Finally, by December 2021, we conducted a thorough disinfection of all surfaces in the ICU, after which no more cases were identified.

In other countries, cases of multidrug resistance were identified in the context of antimicrobial drug pressure and cases of sepsis and pneumonia were diagnosed among immunosuppressed patients (5). In our hospital, 8 (53.3%) patients died. The average time from bacterial isolation to death was 18.2 (range 2–65) days.

*Elizabethkingia* isolates are usually resistant to multiple antibiotics. In analyses of different isolates collected in South Korea and Taiwan (5), all *E. miricola* isolates were resistant to cephalosporins, aminoglycosides, and carbapenems. Those data are similar to results obtained in our hospital (Table). A study conducted in Switzerland found genes encoding metallo- $\beta$ -lactamases in a multidrug-resistant *E. miricola* isolated from the urine of a 2-year-old boy (6). Those genes provide resistance to penicillin- $\beta$ -lactamase inhibitor combinations, carbapenems, cefotaxime, and ceftazidime. Trimethoprim/sulfamethoxazole showed the best antibiogram sensitivity in our outbreak, only 1 of 15 patients showed resistance (Table).

In summary, our study underlines the need to find *Elizabethkingia* spp. bacteria in ICUs. In addition, all species in the *Elizabethkingia* genus should be listed as special surveillance bacteria due to their capacity to cause major illness and death in vulnerable patients. Future studies analyzing differences in the outcomes between patients with *E. miricola* and other patients

admitted to ICU, including patient characteristics and treatments, could expand on the information provided in this study. Finally, to enable early detection of outbreaks of intrinsically antimicrobial-resistant bacteria, modify patient treatment, and save lives, whole-genome sequencing needs to be instituted when rare agents not previously considered for special surveillance are identified.

E.S.-I. and C.V.-U. participated in the treatment and preventive measures for controlling the outbreak. M.R.-I. supervised the work. All authors participated in writing and revising the manuscript for intellectual content.

### About the Author

Dr. Soler-Iborte is a preventive medicine and public health resident at Hospital Universitario San Cecilio, Granada, Spain. Her research interests include nosocomial infections, vaccines, epidemiology and infectious disease prevention in the hospital setting.

### References

- Zdziarski P, Paściak M, Rogala K, Korzeniowska-Kowal A, Gamian A. *Elizabethkingia miricola* as an opportunistic oral pathogen associated with superinfectious complications in humoral immunodeficiency: a case report. *BMC Infect Dis*. 2017;17:763. <https://doi.org/10.1186/s12879-017-2886-7>
- Li Y, Kawamura Y, Fujiwara N, Naka T, Liu H, Huang X, et al. *Chryseobacterium miricola* sp. nov., a novel species isolated from condensation water of space station Mir. *Syst Appl Microbiol*. 2003;26:523–8. <https://doi.org/10.1078/072320203770865828>
- Kim KK, Kim MK, Lim JH, Park HY, Lee ST. Transfer of *Chryseobacterium meningosepticum* and *Chryseobacterium miricola* to *Elizabethkingia* gen. nov. as *Elizabethkingia meningoseptica* comb. nov. and *Elizabethkingia miricola* comb. nov. *Int J Syst Evol Microbiol*. 2005;55:1287–93. <https://doi.org/10.1099/ijs.0.63541-0>
- Gupta P, Zaman K, Mohan B, Taneja N. *Elizabethkingia miricola*: A rare non-fermenter causing urinary tract infection. *World J Clin Cases*. 2017;5:187–90. <https://doi.org/10.12998/wjcc.v5.i5.187>
- Lin JN, Lai CH, Yang CH, Huang YH. *Elizabethkingia* infections in humans: from genomics to clinics. *Microorganisms*. 2019;7:295. <https://doi.org/10.3390/microorganisms7090295>
- Colapietro M, Endimiani A, Sabatini A, Marcoccia F, Celenza G, Segatore B, et al. BlaB-15, a new BlaB metallo- $\beta$ -lactamase variant found in an *Elizabethkingia miricola* clinical isolate. *Diagn Microbiol Infect Dis*. 2016;85:195–7. <https://doi.org/10.1016/j.diagmicrobio.2015.11.016>

Address for correspondence: Eva Soler-Iborte, Service of Preventive Medicine and Public Health, Hospital Universitario San Cecilio, Avenida del Conocimiento s/n 18016, Granada, Spain; email: eva.soler.sspa@juntadeandalucia.es