Chlamydia pneumoniae Upsurge at Tertiary Hospital, Lausanne, Switzerland

Florian Tagini, Onya Opota, Gilbert Greub

Author affiliation: Institute of Microbiology, Lausanne University Hospital, Lausanne, Switzerland

DOI: https://doi.org/10.3201/eid3004.231610

Chlamydia pneumoniae infection cases have usually accounted for <1.5% of community-acquired respiratory tract infections. Currently, Lausanne, Switzerland is experiencing a notable upsurge in cases, with 28 reported within a span of a few months. This upsurge in cases highlights the need for heightened awareness among clinicians.

he intracellular bacterium *Chlamydia pneumoniae* is a recognized cause of community-acquired pneumonia (1). High-frequency estimates were initially derived from serologic studies, but the advent of molecular techniques has revealed rates that are generally <1.5% among patients with respiratory tract infections, although epidemiological change between initial and current rate estimates cannot be ruled out (2,3). Sporadic outbreaks have been documented, such as a 2014 prison outbreak in Texas (4) and a 2016 community-acquired pneumonia outbreak in South Korea (5). In recent years, studies have also linked C. pneumoniae bacteria to bronchitis and asthma (6). C. pneumoniae bacteria has also been documented in patients with cystic fibrosis (7). Of note, infections occur at higher rates in children than in adults (2).

At the height of the SARS-CoV-2 pandemic, *C. pneumoniae* bacteria detection rates were low, paralleling the near-extinction state observed for *Mycoplasma pneumoniae* bacteria in Europe (8). However, a current rebound of *M. pneumoniae* infections is occurring (9). We report a similar increase in PCR-positive *C. pneumoniae* bacteria detection rates at a tertiary hospital in Switzerland. As the case series and the analysis thereof derive from the pathogen surveillance to which our institute is legally bound by the health authorities, Swiss legislation on human research is not applicable and the consent of the patients concerned is not required. This publication complies with the applicable data protection legislation and institutional guidelines.

During routine epidemiologic surveillance at Lausanne University Hospital in Lausanne, Switzerland, positive C. pneumoniae bacteria PCR rates surged to 3.61% during October-December 2023, peaking at 6.66% in October, contrasting with the usual 0%-0.75% range reported over the past decade (Figure 1, panel A, B). The PCR method we used for testing has been previously described in Opota et al. (10). In this most recent outbreak, we documented C. pneumoniae bacteria in 28 patients in 2023; of those, 20 were children (mean age 8 years) and 8 were adults (mean age 43 years). Patients with C. pneumoniae bacteria sometimes reported wheezing as a major clinical complaint. We tested bacterial loads in patients positive for C. pneumoniae bacteria and found that the mean bacterial load was 1,534,821 DNA copies/mL (range 200-11,998,897 DNA copies/mL). We collected nasopharyngeal swabs most frequently (n = 24), whereas we collected sputum samples (n = 5) and nasal swab samples (nostril only, n = 1) less frequently. Of note, bacterial loads were not higher in the analyzed sputa

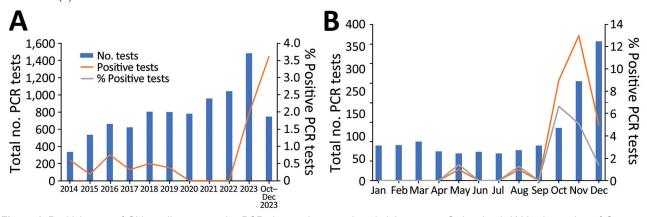


Figure 1. Positivity rate of *Chlamydia pneumoniae* PCRs in a tertiary care hospital, Lausanne, Switzerland. A) Yearly number of *C. pneumoniae* PCR tests conducted during 2014-2023. The final bar shows the last quarter of 2023, when the positivity rate exhibited a notable increase to 3.61%. B) Monthly numbers of *C. pneumoniae* PCR tests performed in 2023, showcasing positive tests and corresponding positivity rates. The data reveal a peak in the percentage of positivity of 6.66% in October.

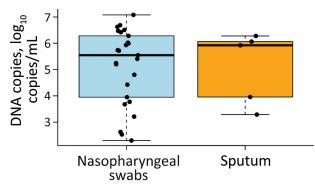


Figure 2. Boxplot of the quantifications of the *Chlamydia pneumoniae*—positive PCRs, by sample type, in a tertiary care hospital, Lausanne, Switzerland. In total, 24 nasopharyngeal swab and 5 sputum samples were available. For 2 patients, data were paired, with 1 nasopharyngeal swab and 1 sputum sample available for each. Black dots indicate individual samples; horizontal lines within boxes indicate medians; box tops and bottoms indicate interquartile range; and error bars indicate 1.5 times the value of the interquartile ranges. The nostril swab was omitted from the analysis. We observed no statistically significant difference between the groups (Wilcoxon rank-sum test).

than in the nasopharyngeal swabs (p = 1 by Wilcoxon rank-sum test) (Figure 2).

The results of this analysis should be interpreted with caution in the absence of a larger number of paired samples. This analysis includes only 2 paired samples exhibiting <1 logarithm (decimal) of difference in DNA copies per milliliter.

To explain this sudden surge of *C. pneumoniae* bacterial infection, we suspect 2 primary factors. First, decreased immunity may have developed because of fewer circulating strains in the population over the past 3 years, related to SARS-CoV-2 transmission prevention measures. Second, recently relaxed hygiene standards after the SARS-CoV-2 pandemic may have increased the risk for infection.

Clinical suspicion of C. pneumoniae infection is particularly warranted when patients' clinical manifestations include a persistent dry cough or wheezing. Molecular testing, if available, should be the first-line diagnostic tool with nasopharyngeal swabs as an acceptable sample collection method. We do not recommend serologic testing in such cases because of the need to collect convalescent serum and the late appearance of antibodies. Antibodies generally develop 2-3 weeks after symptom onset for IgM and 4-8 weeks for IgG, which is rather late for diagnostic and therapeutic purposes. Furthermore, because C. pneumoniae bacterial infection can be treated by macrolides, doxycycline, or fluoroquinolones, current increases in both C. pneumoniae and M. pneumoniae bacteria lead us to

recommend PCR testing for both bacteria in symptomatic patients, instead of testing only for respiratory viruses. Although co-infection with *M. pneumoniae* bacteria occurred in only 1 patient (*M. pneumoniae* PCR was tested on all samples) in our cohort, viral co-infections are not uncommon. Many multiplexed PCR respiratory panels are available and could help monitor the trend of *C. pneumoniae* bacterial infections on a larger scale.

In conclusion, we outline an upsurge of *C. pneumoniae* bacterial infections in the Lausanne region of Switzerland, especially in the pediatric population, raising concerns for other settings and regions. We found no clear epidemiologic link between patients, which suggests that we are detecting a minority of cases and that infections may occur at higher rates in the community than we have documented. This local finding highlights the importance of considering this intracellular bacterium as a causative agent, along with other fastidious organisms such as *M. pneumoniae* bacteria, which are also on the rise (9).

About the Author

Dr. Tagini is a trainee in clinical microbiology and infectious diseases at the Lausanne University Hospital. His research interests are focused mainly on intracellular bacteria and bacterial genomics.

References

- 1. Grayston JT, Campbell LA, Kuo CC, Mordhorst CH, Saikku P, Thom DH, et al. A new respiratory tract pathogen: *Chlamydia pneumoniae* strain TWAR. J Infect Dis. 1990;161:618–25. https://doi.org/10.1093/infdis/161.4.618
- Kumar S, Hammerschlag MR. Acute respiratory infection due to *Chlamydia pneumoniae*: current status of diagnostic methods. Clin Infect Dis. 2007;44:568–76. https://doi.org/ 10.1086/511076
- Senn L, Jaton K, Fitting JW, Greub G. Does respiratory infection due to *Chlamydia pneumoniae* still exist? Clin Infect Dis. 2011;53:847–8. https://doi.org/10.1093/cid/cir515
- Conklin L, Adjemian J, Loo J, Mandal S, Davis C, Parks S, et al. Investigation of a *Chlamydia pneumoniae* outbreak in a federal correctional facility in Texas. Clin Infect Dis. 2013;57:639–47. https://doi.org/10.1093/cid/cit357
- Han HY, Moon JU, Rhim JW, Kang HM, Lee SJ, Yang EA. Surge of *Chlamydia pneumoniae* pneumonia in children hospitalized with community-acquired pneumonia at a single center in Korea in 2016. J Infect Chemother. 2023;29:453–7. https://doi.org/10.1016/j.jiac.2023.01.012
- Hahn DL, Schure A, Patel K, Childs T, Drizik E, Webley W. Chlamydia pneumoniae-specific IgE is prevalent in asthma and is associated with disease severity. PLoS One. 2012;7:e35945. https://doi.org/10.1371/journal.pone. 0035945
- Pittet LF, Bertelli C, Scherz V, Rochat I, Mardegan C, Brouillet R, et al. *Chlamydia pneumoniae* and *Mycoplasma pneumoniae* in children with cystic fibrosis: impact on bacterial respiratory microbiota diversity. Pathog Dis. 2021;79:ftaa074.

RESEARCH LETTERS

- Meyer Sauteur PM, Beeton ML, Pereyre S, Bébéar C, Gardette M, Hénin N, et al.; ESGMAC the ESGMAC MAPS study group. *Mycoplasma pneumoniae*: gone forever? Lancet Microbe. 2023;4:e763. https://doi.org/10.1016/ S2666-5247(23)00182-9
- Meyer Sauteur PM, Beeton ML, Pereyre S, Bébéar C, Gardette M, Hénin N, et al. European Society of Clinical Microbiology and Infectious Diseases (ESCMID) Study Group for Mycoplasma and Chlamydia Infections (ESGMAC), and the ESGMAC Mycoplasma pneumoniae Surveillance (MAPS) Study Group. Mycoplasma pneumoniae: delayed re-emergence after COVID-19 pandemic restrictions. Lancet Microbe. 2024;5:e100-1. https://doi.org/10.1016/ S2666-5247(23)00344-0
- Opota O, Brouillet R, Greub G, Jaton K. Methods for realtime PCR-based diagnosis of *Chlamydia pneumoniae*, *Chlamydia psittaci*, and *Chlamydia abortus* infections in an opened molecular diagnostic platform. Methods Mol Biol. 2017;1616:171–81. https://doi.org/10.1007/978-1-4939-7037-7_11

Address for correspondence: Gilbert Greub, Institute of Microbiology, Department of Laboratory Medicine and Pathology, Lausanne University Hospital and University of Lausanne, Bugnon 48, CH-1011 Lausanne, Switzerland; email: gilbert.greub@chuv.ch

Highly Pathogenic Avian Influenza A(H5N1) Viruses from Multispecies Outbreak, Argentina, August 2023

Agustina Rimondi, Ralph E.T. Vanstreels, Valeria Olivera, Agustina Donini, Martina Miqueo Lauriente, Marcela M. Uhart

Author affiliations: Robert Koch Institute, Berlin, Germany (A. Rimondi); Instituto Nacional de Tecnología Agropecuaria Instituto de Virología e Innovaciones Tecnológicas, Buenos Aires, Argentina (A. Rimondi, V. Olivera); University of California School of Veterinary Medicine, Davis, California, USA (R.E.T. Vanstreels, M.M. Uhart); Southern Right Whale Health Monitoring Program, Puerto Madryn, Argentina (A. Donini, M.M. Uhart); Secretaria de Ambiente y Cambio Climático de Provincia de Río Negro, Viedma, Argentina (M. Miqueo Lauriente)

DOI: https://doi.org/10.3201/eid3004.231725

We report full-genome characterization of highly pathogenic avian influenza A(H5N1) clade 2.3.4.4b virus from an outbreak among sea lions (August 2023) in Argentina and possible spillover to fur seals and terns. Mammalian adaptation mutations in virus isolated from marine mammals and a human in Chile were detected in mammalian and avian hosts.

In February 2023, the first case of highly pathogenic **L**avian influenza (HPAI) A(H5N1) in Argentina was detected in a wild goose near the border with Bolivia and Chile (Appendix Figure 1, https://wwwnc. cdc.gov/EID/article/30/4/23-1725-App1.pdf) (1). In contrast with Peru and Chile, where extensive mortality of seabirds and marine mammals had been attributed to the virus in the preceding months (2,3), the initial spread of HPAI H5N1 in Argentina was largely limited to backyard and industrial poultry (94 outbreaks), causing the death or disposal of 2.2 million birds. Argentina declared itself free from the disease in poultry on August 8, 2023; before then, HPAI H5N1 detections in wildlife in Argentina had been scarce (7 events during February-April) and limited to aquatic birds (Anatidae, Laridae, and Rallidae) (1,4). However, soon thereafter, the national animal health services confirmed HPAI H5N1 in South American sea lions (Otaria byronia) from Río Grande, southernmost Argentina. Over subsequent weeks, the virus was detected in sea lions northward along the Argentina coast, and sporadic cases also occurred in South American fur seals (Arctocephalus australis). The most affected site was Punta Bermeja (Appendix Figure 1), the largest sea lion colony in Argentina, where an estimated 811 sea lions died over 2 months; minimal numbers (<5) of fur seals and terns were also affected (1,4).

In collaboration with provincial authorities and park rangers, we collected swab samples (oronasal, rectal, tracheal, lung, and brain) from 16 deceased sea lions, 1 fur seal, 1 great grebe (*Podiceps major*), and 1 South American tern (Sterna hirundinacea) discovered at Punta Bermeja on August 26, 2023. A sampled adult male sea lion was seen alive showing clinical signs consistent with HPAI infection (inability to stand or walk, muscular tremors and spasms, difficulty breathing, and abundant oral mucus). We tested the samples by real-time reverse transcription PCR targeting influenza A virus (5) and confirmed that all were positive. On the basis of viral RNA yields, we selected brain samples from 4 sea lions, 1 fur seal, and 1 tern for full-genome sequencing (Appendix Figure 2). We used maximum-likelihood tree phylogenetic analysis (6) and mutational analysis